

Research



Cite this article: Stockmaier S, Bolnick DI, Page RA, Josic D, Carter GG. 2020 Immune-challenged vampire bats produce fewer contact calls. *Biol. Lett.* **16**: 20200272.

<http://dx.doi.org/10.1098/rsbl.2020.0272>

Received: 21 April 2020

Accepted: 23 June 2020

Subject Areas:

behaviour, health and disease and epidemiology

Keywords:

sickness behaviour, pathogen transmission, infection, social behaviour, lipopolysaccharide

Author for correspondence:

Sebastian Stockmaier

e-mail: sebastian.stockmaier@utexas.edu

Electronic supplementary material is available online at <https://doi.org/10.6084/m9.figshare.c.5046795>.

Animal behaviour

Immune-challenged vampire bats produce fewer contact calls

Sebastian Stockmaier^{1,2}, Daniel I. Bolnick^{1,3}, Rachel A. Page², Darija Josic⁴ and Gerald G. Carter^{2,5}

¹Department of Integrative Biology, University of Texas at Austin, Austin, TX 78712, USA

²Smithsonian Tropical Research Institute, Apartado Postal 0843-03092, Balboa, Ancon, Panama

³Department of Ecology and Evolutionary Biology, University of Connecticut, Storrs, CT 06269-3043, USA

⁴Museum fuer Naturkunde, Leibniz-Institute for Research on Evolution and Biodiversity, 10115 Berlin, Germany

⁵Department of Evolution, Ecology and Organismal Biology, The Ohio State University, Columbus, OH 43210, USA

SS, 0000-0001-8280-8086; DIB, 0000-0003-3148-6296; RAP, 0000-0001-7072-0669; GGC, 0000-0001-6933-5501

Vocalizations are an important means to facilitate social interactions, but vocal communication may be affected by infections. While such effects have been shown for mate-attraction calls, other vocalizations that facilitate social contact have received less attention. When isolated, vampire bats produce contact calls that attract highly associated groupmates. Here, we test the effect of an immune challenge on contact calling rates of individually isolated vampire bats. Sickness behaviour did not appear to change call structure, but it decreased the number of contact calls produced. This effect could decrease contact with groupmates and augment other established mechanisms by which sickness reduces social encounters (e.g. mortality, lethargy and social withdrawal or disinterest).

1. Introduction

Infections can reduce contact between individuals by inducing ‘sickness behavior’. For instance, sickness can decrease physical social encounters through reduced movement [1] or decrease directed social interactions like grooming [2,3]. Reductions in social contact can also occur if infected individuals vocalize less. For example, an immune challenge reduces male mate-attraction vocalizations or some of their components in several species [4–8]. If sick males attract fewer females and if avoiding sick males decreases the likelihood of females acquiring parasites [9–11], then transmission of parasites between the sexes will decrease [12].

Besides courtship vocalizations, a broader range of vocal interactions could be influenced by sickness, but these other call types have received less attention. In many group-living animals that live in conditions of low visibility, or that must maintain cohesion while on the move, individuals produce contact calls to maintain contact with groupmates or particular affiliated individuals [13–21]. If contact calls facilitate physical contact, then sickness behaviour that reduces the rate of contact calling should decrease contact with groupmates. However, if contact calling is used by an individual in need to gain benefits from others, then sick individuals might instead make a greater number of contact calls. For example, when parents can acquire enough food to feed all their offspring, hungry nestlings in worse condition are expected to call more often, not less [22,23]. The expected effect of sickness behaviour on contact calling by distressed individuals is therefore less clear.

Isolated common vampire bats (*Desmodus rotundus*) produce multi-harmonic contact calls that vary in spectral structure and duration, and that facilitate individual contact and recognition ([15,17], figure 1*b,c*). Contact calls appear to be

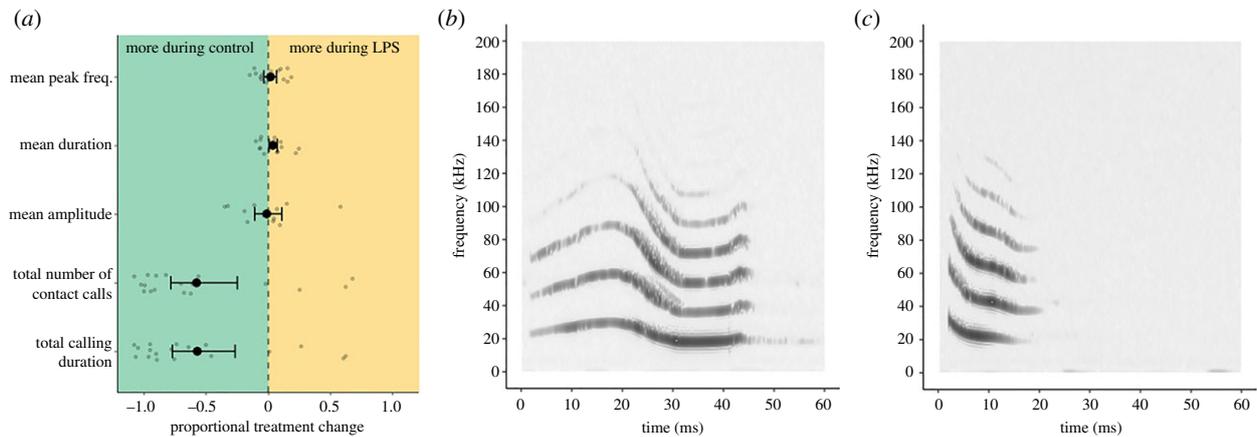


Figure 1. Effect of lipopolysaccharide (LPS) on vampire bat contact calls. (a) Shows mean standardized LPS effect \pm bootstrapped 95% confidence intervals (see electronic supplementary material, table S1) and data points for the mean peak frequency, mean duration, mean amplitude, the total number of contact calls produced and the total calling duration. Example spectrograms of a longer (b) and shorter (c) vampire bat contact call that also vary in peak frequency (darker regions show frequencies of greater relative amplitude).

important for maintaining co-roosting associations with bonded partners and for finding or recruiting those partners for help; for example, trapped and hungry individuals appear to use contact calls to recruit both kin and non-kin food donors to feed them by regurgitation [17,24].

Here, we mimicked a bacterial infection in vampire bats using lipopolysaccharide (LPS) to trigger transient physiological symptoms and sickness behaviours [2,3,25], and then we tested for the effect of LPS on contact calling behaviour. LPS-injected vampire bats are groomed by fewer bats and have lower social connectedness in the wild, an effect that could be driven in part by a reduction in contact calling [3,25]. We show that LPS-induced sickness behaviour decreases the number of contact calls produced by isolated vampire bats. This effect is relevant for pathogen transmission in social animals that rely on vocalizations to maintain contact because it might further reduce the probability of physical contact between individuals beyond the effects of reduced movement.

2. Material and methods

We recorded contact calls by physically isolating an adult female vampire bat ($n = 18$) in a soft mesh cage at a distance of 10–30 cm from a CM16 ultrasound condenser microphone (frequency range 1–200 KHz, Avisoft Bioacoustics, Berlin, Germany). The mesh cage was inside a 68 l plastic bin lined with acoustic dampening foam and within hearing range of conspecifics of a captive colony. To selectively record contact calls, we used a digitizer (116 Hn UltrasoundGate, Avisoft Bioacoustics, Berlin, Germany, sampling frequency of 250 or 500 kHz) to save a .wav file whenever a 10–50 kHz sound was detected at greater than 5% amplitude. We used Avisoft SASLabPro (Avisoft Bioacoustics, Berlin, Germany) to measure the onset, duration, peak amplitude and peak frequency of all calls. We excluded echolocation calls and other noise by deleting sounds that were longer than 60 ms and shorter than 10 ms.

For each bat, we recorded two trial types. In LPS trials, we induced sickness behaviour in subjects by injecting them subcutaneously with LPS (L2630 Sigma-Aldrich, USA, dose: 5 mg kg⁻¹ body mass of bat) in phosphate-buffered saline (PBS) before the recording period. We chose this dose based initially on observed effects in another bat species [26], and on later studies in vampire bats, which showed that this dose increases white blood cell count and neutrophil to lymphocyte ratio [2] and decreases

physical activity, social encounters and social grooming [2,3,25]. In control trials, the same bats were injected with an equivalent volume of only PBS as a control treatment.

Treatments were given in random order, and eight bats received the control treatment first. We recorded bats for 4–6 h immediately after the injection, because we previously detected symptoms for at least 6 h post-injection [2]. Different bats were recorded for different times after injection, but the paired LPS and control trials were always the same duration and time of the night. Since bats often sleep in the recording chamber, we excluded hours when the bat did not call during either the treatment night or control night, but our results do not change if we include these hours in our analysis. The inter-trial period was at least 5 days to ensure recovery of the bats [2,25]. To calculate a standardized effect (proportional change) of LPS on vocalizations for each bat, we used $(Y_{LPS} - Y_C)/(Y_{LPS} + Y_C)$, where Y_{LPS} and Y_C are the measures of vocal activity during the bat's LPS and control trial, respectively.

To test for an effect of LPS on contact calling, we randomly swapped the control and LPS trial data within each bat to calculate a distribution of t -statistics under the null hypothesis of no difference between the LPS and control trial, then compared the observed t -statistic to this distribution to obtain a two-sided p -value (i.e. a nonparametric permuted paired t -test). To estimate 95% confidence intervals for LPS effect sizes, we used nonparametric bootstrapping with accelerated bias-corrected percentile limits [27]. We used 5000 permutations for both methods. We calculated the mean and bootstrapped 95% CI for the LPS effect on five measures of contact calling behaviour: the total number of contact calls produced, the sum of call durations, the mean call duration, the mean amplitude and the mean peak frequency (the frequency at the point of the maximum amplitude of the entire element). Data and R script to repeat our analysis are available on figshare [28].

3. Results

LPS injections led to fewer contact calls. The average contact calling rates per bat during the control and LPS trials were, respectively, 66 and 16 contact calls per hour (see electronic supplementary material, figure S1 for details on each bat and electronic supplementary material, figure S2 for average call production of bats after LPS and control over time). On average, LPS injections caused female vampire bats to produce 30% fewer contact calls, with 15 of 18 bats producing fewer contact calls during the LPS trial compared to the

Table 1. Effect of LPS on vampire bat contact calls. Means and their bootstrapped 95% confidence intervals for the standardized LPS effects for contact call number, total calling duration, mean amplitude, mean peak frequency and mean call duration (table corresponds to figure 1a).

measure	N	lower CI	mean	upper CI	p-value
call number	18	−0.78	−0.58	−0.27	0.0006
calling duration	18	−0.77	−0.57	−0.26	0.0012
mean amplitude	14 ^a	−0.11	−0.01	0.11	0.8754
mean peak frequency	14 ^a	−0.03	0.02	0.07	0.4656
mean duration	14 ^a	0.01	0.04	0.08	0.0510

^aWe could not calculate the LPS effect on mean amplitude, mean peak frequency and mean duration for four bats because they did not produce any calls when injected with LPS.

control trial ($p = 0.0006$, figure 1a and table 1, electronic supplementary material, figure S1). Fewer calls led to an average decrease of 32% in total calling duration ($p = 0.0012$, figure 1a and table 1). We did not detect an effect of LPS on mean call amplitude (figure 1a and table 1). Although vampire bats produce contact calls that vary in call structure (figure 1b,c), we did not detect an effect of LPS injections on mean call duration (figure 1a and table 1) or mean peak frequency (figure 1a and table 1).

4. Discussion

Infection-induced sickness behaviours can affect vocal communication as evident in LPS-injected male house mice that produce fewer call syllables, which likely contributes to reduced associations with females [7]. Similarly, immune-challenged males decrease their song rate in collared flycatchers (*Ficedula albicollis*), white-browed sparrow weavers (*Plocepasser mahali*), field crickets (*Gryllus campestris*) and white-crowned sparrows (*Zonotrichia leucophrys gambelii*) [5,6,8,29]. In comparison to these mate-attraction calls, contact calls and signals of need are interesting to consider because a state of poor condition could lead to either a higher or lower calling rate. In vampire bats, contact calling can attract food donors and might act as a signal of need [15,17]. Here, we showed that an immune challenge reduces contact calling, which could potentially help to explain why immune-challenged vampire bats encounter fewer individuals [3,25], in addition to the more obvious explanation of reduced movement.

In a previous study using the same dose as we used here, LPS-injected vampire bats were more lethargic, spending less time awake, moving or engaging in hygienic behaviours such as self-grooming [2]. So, our results are most consistent with the simplest explanation that reduced contact calling is also owing to lethargy. Reduced contact calling is unlikely to be explained as a kin-selected mechanism for reducing pathogen transmission [30–32] because contact calls attract both kin and non-kin [17]. We hypothesize that vampire bats reduce contact calling to support the energetic demands of the physiological response. Across several taxa, the metabolic costs of acoustic signalling are estimated to be about eight times that of remaining silent [33] and call rate is sensitive to other ecological constraints like reduced food availability [34]. Experiments with a related frugivorous bat species show that LPS injections reduce body mass and increase resting metabolic rate by 40% [35].

We used a dose of LPS for which we knew the physiological and behavioural effects in vampire bats [2,3,25]. It is important to note, however, that the physiological responses to LPS are dose-dependent and involve both pro-inflammatory and anti-inflammatory responses [36]. To determine what doses are most ecologically relevant for different diseases, future work must compare the relationship between the dose-dependent effects of LPS against the effects of natural bacterial infections in vampire bats and other species.

It is also important to note that infection-induced changes to social vocalizations are pathogen specific. LPS mimics common symptoms of a bacterial infection in vampire bats and other animals [1,2]. Some live pathogens, however, could increase specific social behaviours to favour their transmission [37]. For instance, chytrid-fungus infected Japanese tree frogs (*Hyla japonica*) increase their mating call effort, which potentially favours the transmission of the fungus [38]. Vampire bats harbour multiple pathogens in their saliva that rely on directed social interactions, like *Bartonella* [39], hemoplasmas [40] and, most notably, rabies [41]. It would be particularly interesting to look at how rabid vampire bats change their calling behaviour and their response to calls of conspecifics.

Besides call rate, the structure of animal vocalizations might also depend on infection status [4,42]. Common vampire bats produce highly variable contact calls (e.g. figure 1b and c), but we found no evidence that sick bats consistently produced any particular contact call structure more or less. However, for some pathogens, such as rabies, which could affect the vocal tract, there may be clear differences in call structure.

Ethics. Our work was approved by the Smithsonian Tropical Research Institute Animal Care and Use Committee (#2016-0728-2019-A2), the Animal Care and Use Committee of the University of Texas at Austin (AUP-2016-00124) and by the Panamanian Ministry of Environment (protocol: SE/A-64-17).

Data accessibility. We have uploaded Data and R code to a figshare repository. The repository is cited in the methods part of the main text: Stockmaier S. 2020 Dataset and R.code from: 'Sickness behavior reduces contact calling in vampire bats'. Figshare digital repository. <https://doi.org/10.6084/m9.figshare.11861877.v4>.

Authors' contributions. S.S. and G.G.C. designed the study and carried out the experiments and husbandry. G.G.C. captured the bats and established the captive colony. S.S., G.G.C. and D.J. contributed to the data collection and analysis. G.G.C., D.I.B. and R.A.P. coordinated the study and provided valuable resources and laboratory space. All authors contributed to draft the manuscript, gave final approval for

publication and agree to be held accountable for the work performed therein.

Competing interests. We declare we have no competing interests.

Funding. This study was supported by Smithsonian Tropical Research Institute.

Acknowledgements. We thank the Smithsonian Tropical Research Institute for logistical support. Samuel Kaiser, Vanessa Pérez, Jineth Berrio-Martinez, Imran Razik, Bridget Brown, David Girbino, Simon Ripperger and Emma Kline for help in the field and animal caretaking.

References

- Lopes PC, Block P, König B. 2016 Infection-induced behavioural changes reduce connectivity and the potential for disease spread in wild mice contact networks. *Sci. Rep.* **6**, 31790. (doi:10.1038/srep31790)
- Stockmaier S, Bolnick DI, Page RA, Carter GG. 2018 An immune challenge reduces social grooming in vampire bats. *Anim. Behav.* **140**, 141–149. (doi:10.1016/j.anbehav.2018.04.021)
- Stockmaier S, Bolnick DI, Page RA, Carter GG. 2020 Sickness effects on social interactions depend on the type of behaviour and relationship. *J. Anim. Ecol.* **89**, 1387–1394. (doi:10.1111/1365-2656.13193)
- Dreiss AN, Navarro C, De Lope F, Møller AP. 2008 Effects of an immune challenge on multiple components of song display in barn swallows *Hirundo rustica*: implications for sexual selection. *Ethology* **114**, 955–964. (doi:10.1111/j.1439-0310.2008.01546.x)
- Garamszegi LZ, Møller AP, Török J, Michl G, Péczely P, Richard M. 2004 Immune challenge mediates vocal communication in a passerine bird: an experiment. *Behav. Ecol.* **15**, 148–157. (doi:10.1093/beheco/arg108)
- Jacot A, Scheuber H, Brinkhof MWG. 2004 Costs of an induced immune response on sexual display and longevity in field crickets. *Evolution* **58**, 2280–2286. (doi:10.1111/j.0014-3820.2004.tb01603.x)
- Lopes PC, König B. 2016 Choosing a healthy mate: sexually attractive traits as reliable indicators of current disease status in house mice. *Anim. Behav.* **111**, 119–126. (doi:10.1016/j.anbehav.2015.10.011)
- York JE, Radford AN, Grootuis TG, Young AJ. 2016 Dominant male song performance reflects current immune state in a cooperatively breeding songbird. *Ecol. Evol.* **6**, 1008–1015. (doi:10.1002/ece3.1938)
- Hillgarth N. 1996 Ectoparasite transfer during mating in ring-necked pheasants *Phasianus colchicus*. *J. Avian Biol.* **27**, 260–262. (doi:10.2307/3677232)
- Luong LT, Platzer EG, Zuk M, Giblin-Davis RM. 2000 Venereal worms: sexually transmitted nematodes in the decorated cricket. *J. Parasitol.* **86**, 471–477. (doi:10.1645/0022-3395(2000)086[0471:VWSTNI]2.0.CO;2)
- Martinez-Padilla J, Vergara P, Mougeot F, Redpath SM. 2012 Parasitized mates increase infection risk for partners. *Am. Nat.* **179**, 811–820. (doi:10.1086/665664)
- Able DJ. 1996 The contagion indicator hypothesis for parasite-mediated sexual selection. *Proc. Natl Acad. Sci. USA* **93**, 2229–2233. (doi:10.1073/pnas.93.5.2229)
- Arnold BD, Wilkinson GS. 2011 Individual specific contact calls of pallid bats (*Antrozous pallidus*) attract conspecifics at roosting sites. *Behav. Ecol. Sociobiol.* **65**, 1581–1593. (doi:10.1007/s00265-011-1168-4)
- Bradbury JW, Vehrencamp SL. 1998 *Principles of animal communication*. Sunderland, MA: Sinauer Associates.
- Carter GG, Logsdon R, Arnold BD, Menchaca A, Medellín RA. 2012 Adult vampire bats produce contact calls when isolated: acoustic variation by species, population, colony, and individual. *PLoS ONE* **7**, e38791. (doi:10.1371/journal.pone.0038791)
- Carter GG, Fenton MB, Faure PA. 2009 White-winged vampire bats (*Diaemus youngi*) exchange contact calls. *Can. J. Zool.* **87**, 604–608. (doi:10.1139/Z09-051)
- Carter GG, Wilkinson GS. 2016 Common vampire bat contact calls attract past food-sharing partners. *Anim. Behav.* **116**, 45–51. (doi:10.1016/j.anbehav.2016.03.005)
- Cortopassi KA, Bradbury JW. 2006 Contact call diversity in wild orange-fronted parakeet pairs, *Aratinga canicularis*. *Anim. Behav.* **71**, 1141–1154. (doi:10.1016/j.anbehav.2005.09.011)
- Janik VM, Sayigh LS. 2013 Communication in bottlenose dolphins: 50 years of signature whistle research. *J. Comp. Physiol. A Neuroethol. Sens. Neural. Behav. Physiol.* **199**, 479–489. (doi:10.1007/s00359-013-0817-7)
- Maurello MA, Clarke JA, Ackley RS. 2000 Signature characteristics in contact calls of the white-nosed Coati. *J. Mammal.* **81**, 415–421. (doi:10.1644/1545-1542(2000)081<0415:SCICO>2.0.CO;2)
- van Oosterom L, Montgomery JC, Jeffs AG, Radford CA. 2016 Evidence for contact calls in fish: conspecific vocalisations and ambient soundscape influence group cohesion in a nocturnal species. *Sci. Rep.* **6**, 19098. (doi:10.1038/srep19098)
- Caro SM, Griffin AS, Hinde CA, West SA. 2016 Unpredictable environments lead to the evolution of parental neglect in birds. *Nat. Commun.* **7**, 10985. (doi:10.1038/ncomms10985)
- Godfray HJ. 1991 Signalling of need by offspring to their parents. *Nature* **352**, 328–330. (doi:10.1038/352328a0)
- Carter GG, Wilkinson GS, Page RA. 2017 Food-sharing vampire bats are more nepotistic under conditions of perceived risk. *Behav. Ecol.* **28**, 565–569. (doi:10.1093/beheco/ax006)
- Ripperger SP, Stockmaier S, Carter GG. 2020 Sickness behaviour reduces network centrality in wild vampire bats. *bioRxiv* 2020.03.30.015545. (doi:10.1101/2020.03.30.015545)
- Stockmaier S, Dechmann DK, Page RA, O'Mara MT. 2015 No fever and leucocytosis in response to a lipopolysaccharide challenge in an insectivorous bat. *Biol. Lett.* **11**, 20150576. (doi:10.1098/rsbl.2015.0576)
- Puth M-T, Neuhäuser M, Ruxton GD. 2015 On the variety of methods for calculating confidence intervals by bootstrapping. *J. Anim. Ecol.* **84**, 892–897. (doi:10.1111/1365-2656.12382)
- Stockmaier S. 2020 Dataset and R.code from: 'Sickness behavior reduces contact calling in vampire bats'. Figshare digital repository. (doi:10.6084/m9.figshare.11861877.v4)
- Owen-Ashley NT, Turner M, Hahn TP, Wingfield JC. 2006 Hormonal, behavioral, and thermoregulatory responses to bacterial lipopolysaccharide in captive and free-living white-crowned sparrows (*Zonotrichia leucophrys gambelii*). *Horm. Behav.* **49**, 15–29. (doi:10.1016/j.yhbeh.2005.04.009)
- Bos N, Lefevre T, Jensen AB, d'Ettorre P. 2012 Sick ants become unsociable. *J. Evol. Biol.* **25**, 342–351. (doi:10.1111/j.1420-9101.2011.02425.x)
- Heinze J, Walter B. 2010 Moribund ants leave their nests to die in social isolation. *Curr. Biol.* **20**, 249–252. (doi:10.1016/j.cub.2009.12.031)
- Stroeymeyt N, Grasse AV, Crespi A, Mersch DP, Cremer S, Keller L. 2018 Social network plasticity decreases disease transmission in a eusocial insect. *Science* **362**, 941–945. (doi:10.1126/science.aat4793)
- Ophir AG, Schrader SB, Gillooly JF. 2010 Energetic cost of calling: general constraints and species-specific differences. *J. Evol. Biol.* **23**, 1564–1569. (doi:10.1111/j.1420-9101.2010.02005.x)
- Ritschard M, Brumm H. 2012 Zebra finch song reflects current food availability. *Evol. Ecol.* **26**, 801–812. (doi:10.1007/s10682-011-9541-3)
- Guerrero A, Rivera D, Díaz V, Triana C, Nino A. 2018 Metabolic cost of acute phase response in the frugivorous bat, *Artibeus lituratus*. *Mamm. Res.* **63**, 397–404. (doi:10.1007/s13364-018-0375-z)
- Armour EM, Bruner TL, Hines JK, Butler MW. 2020 Low-dose immune challenges result in detectable levels of oxidative damage. *J. Exp. Biol.* **223**, jeb220095. (doi:10.1242/jeb.220095)
- Klein SL. 2003 Parasite manipulation of the proximate mechanisms that mediate social behavior in vertebrates. *Tribute Paul MacLean Neurobiol. Relev. Soc. Behav.* **79**, 441–449. (doi:10.1016/S0031-9384(03)00163-X)
- An D, Waldman B. 2016 Enhanced call effort in Japanese tree frogs infected by amphibian chytrid

- fungus. *Biol. Lett.* **12**, 20160018. (doi:10.1098/rsbl.2016.0018)
39. Becker DJ, Bergner LM, Bentz AB, Orton RJ, Altizer S, Streicker DG. 2018 Genetic diversity, infection prevalence, and possible transmission routes of *Bartonella* spp. in vampire bats. *PLoS Negl. Trop. Dis.* **12**, e0006786. (doi:10.1371/journal.pntd.0006786)
40. Volokhov DV, Becker DJ, Bergner LM, Camus MS, Orton RJ, Chizhikov VE, Altizer SM, Streicker DG. 2017 Novel hemotropic mycoplasmas are widespread and genetically diverse in vampire bats. *Epidemiol. Infect.* **145**, 3154–3167. (doi:10.1017/S095026881700231X)
41. Aguilar-Setien A, Loza-Rubio E, Salas-Rojas M, Brisseau N, Cliquet F, Pastoret PP, Rojas-Dotor S, Tesoro E, Kretschmer R. 2005 Salivary excretion of rabies virus by healthy vampire bats. *Epidemiol. Infect.* **133**, 517–522. (doi:10.1017/S0950268805003705)
42. Fedorka KM, Mousseau TA. 2006 Immune system activation affects male sexual signal and reproductive potential in crickets. *Behav. Ecol.* **18**, 231–235. (doi:10.1093/beheco/arl067)